

## PREPARING NANOPARTICLES USING SOME BIOLOGICAL METHODS AND STUDYING THEIR EFFECT ON SOME MYCOTOXINS

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**Abstract: General Background:** Mycotoxins, toxic substances produced by certain fungi and bacteria, pose significant health risks to humans and animals, necessitating effective remediation strategies. **Specific Background:** The use of environmentally friendly approaches to mitigate the impact of these toxins is crucial. Nanotechnology, particularly in conjunction with plant extracts, has emerged as a promising method for toxin remediation due to its cost-effectiveness and efficiency. **Knowledge Gap:** Despite the potential of nanomaterials in mycotoxin management, there is limited research exploring the interaction between plant extracts and nanotechnology in inhibiting toxin-producing fungi. **Aims:** This study aims to investigate the efficacy of zinc oxide nanomaterials synthesized from mint plant extracts in mitigating the effects of mycotoxins and inhibiting the growth of mycotoxin-producing fungi. **Results:** Mint leaves were collected from various locations in Al-Qadisiyah Governorate over an eight-week period. The extracted plant materials were used to synthesize zinc oxide nanoparticles, which demonstrated significant inhibitory effects on the growth of specific fungi responsible for mycotoxin production, thereby reducing mycotoxin formation. **Novelty:** This research highlights the innovative application of mint-derived zinc oxide nanoparticles as a dual-action agent—suppressing fungal growth while simultaneously preventing mycotoxin synthesis. **Implications:** The findings provide valuable insights into sustainable agricultural practices and mycotoxin management, paving the way for the development of natural, eco-friendly solutions to enhance food safety and protect public health.

**Keywords:** Mycotoxins, Nanotechnology, Zinc Oxide, Plant Extract, Fungal Inhibition, Al-Qadisiyah.



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### Introduction

Mycotoxins are toxic by-products produced by some species of filamentous fungi and have long-term negative effects for humans and animals alike. Although therapeutic methods are constantly improving, the best method is nanotechnology, which has been used in all fields due to its low cost. Its effectiveness in eliminating fungal toxins (Horky et al., 2018). Some types of fungi secrete some secondary metabolic substances that may have a harmful effect on animals and humans. These secondary substances are called mycotoxins (Turner et al., 2010). In view of most of the old methods of inhibiting the growth of fungi, researchers have been proposing new ideas and methods to inhibit the growth of fungi and mycotoxins by using nanoparticles (Jogee & Rai, 2020). Mycotoxins have serious effects on health (such as cancer, birth defects, and genetic mutations), and this represents a

global concern, To ensure human health, mycotoxins must be monitored, because in recent years there have been many poisoning incidents due to mycotoxins (Yang et al., 2020). The most important types of fungi that produce mycotoxins are *Aspergillus* species and *Fusarium* species. Includes *Aspergillus* {ochratoxin A (OTA), aflatoxins (AFL), trichothecenes and deoxynivalenols (DON)}, The most important mycotoxins are aflatoxins, and the most important types of these toxins are B1, B2, G1, and G2. It is produced by two types of fungi, *Aspergillus parasiticus* and *Aspergillus flavus*, which are abundant in grains, soil, and decaying plants (Elkenany & Awad., 2021). They are classified according to the value of the flow rate (rF) on thin layer chromatography (tlc) slides, as well as according to the color of their fluorescence under ultraviolet rays (Qi et al., 2015). but *Fusarium* includes {fumonisins (FUM) B1 and B2, zearalenone (ZEA)} and the emerging mycotoxins fusaproliferin., moniliformin, pefercin, Eniatin), *Claviceps* (ergot alkaloids), and *Alternaria* (alternariol, alternariol, alternariol methyl ether, altertoxin, and tenosonic acid) (Cunha et al., 2018). For many years, the mechanism of action of mycotoxins and their dangerous effects have been studied in terms of disorders of the central nervous system, poisoning of the heart, liver, and digestive system, as well as kidney poisoning (Sobral et al., 2018; Dellaflora et al., 2018; Freire and Sant'Ana., 2018). Mycotoxins have a toxic effect on animal species. Poultry is considered the most sensitive, then pigs and ruminants (Cheat et al., 2016). With almost 100% efficiency, ruminants metabolize some types of mycotoxins (Rodrigues., 2014). Because mycotoxins are lipophilic, they are preferred to be part of animal products (Krstanovic et al., 2017). For example, AFL M1 toxin easily enters milk, which may cause significant health effects (Aslam et al., 2015). As a result of food contaminated with these mycotoxins, it may lead to acute or chronic poisoning of animals and humans (El-Sayed et al., 2022).

Nanotechnology offers many solutions to control many pathogens as it has environmentally friendly advantages (Gacem et al., 2020). Many nanomaterials have been used to treat mycotoxins through the rapid development of this type of technology (Jiang et al., 2018). In the past, there has been an increase in nanotechnology due to its use in several applications such as medicine, chemistry, etc (Pelaz et al., 2012). Most recent studies show that the green synthesis of metals is of high importance, as metal oxides such as zinc, copper, silver, gold, and nickel are gaining ground (Moodley et al., 2018; Suresh et al., 2018). Among various metal oxides, ZnO nanoparticles exhibit high electronic mobility, large exciton binding energy, wide band gap, and high optical transmittance (Kołodziejczak-Radzimska & Jesionowski., 2014). Metal oxides and nanostructures exhibit chemical and biophysical properties that include extraordinary optical properties, catalytic activity, thermomechanical properties, and antimicrobial activity against pathogenic microbes (Geethalakshmi & Sarada., 2013; Aiad et al., 2014). Nanoparticles acquire multiple properties due to their small size, which makes them desirable for many uses, such as cosmetics (Raj et al., 2012). Recently, it has been observed that there is great interest around the world in the antifungal activity of zinc oxide nanoparticles (ZnO NPs), due to the emergence of severe drug resistance by microorganisms. Nanomaterials have clearly shown high activity against multidrug-resistant microorganisms (Ashour et al., 2020; El-Tarabily et al., 2021). Most types of food, after being exposed to contamination or unsuitable conditions, contain fungi capable of producing a toxic substance or a group of toxins that are harmful to human and animal health (ElHamaky et al., 2016).

The mechanism of action of the chemicals present in the plant extract is as a reducing agent to convert mineral precursors into nanoparticles. These materials are characterized by being non-toxic and acting as antioxidants (Singh et al., 2018). The most important phytochemicals that contribute to the reduction process are aldehydes, alkaloids, and phenolic compounds. Also, the concentration of

mineral salts, pH, and temperature affect (stability, quantity, and synthesis of nanoparticles) (Mukunthan & Balaji.,2012).

The aim of this study is to control fungi that secrete mycotoxins using environmentally friendly nanoparticles.

## **Methods**

### **2.1- Collect plant samples**

Mint plants were collected from some grocery stores and gardens located in Al-Qadisiyah Governorate and from various places (Al-Diwaniyah, Afak District, Al-Shamiya District, Al-Shinafiya District, Al-Daghara District, and Al-Nouriya District) on 2/3/2024 to 3/29/2024.

### **2.2-Preparation of plant extract from mint leaves**

The plant extract is first prepared by collecting the leaves of the plant used for study and removing all dust and particles stuck in it by washing with distilled water, then drying it at room temperature, Then we weigh 100 grams of it, cut it into small parts, add a little water to it and grind it using a mortar and pestle to soften the cell membrane. The process is done the extract is boiled for 15 minutes and in order to remove the broken cell parts, it is filtered with filter paper, then we use a centrifuge at 2400 rpm for 5 minutes, then we use the pure extract to synthesize the required nanoparticles (Jayachandran et al.,2021).

### **2.3-Synthesis of zinc oxide nanoparticles**

Zinc nitrate was added to the previously prepared plant extract and for 20 minutes, the mixture was stirred at a temperature of 65 degrees Celsius. Then we began to notice the color changing from green to light yellow. After that, it was left at the same temperature overnight until we obtained a thick yellow paste. It was left to dry and calcined at a temperature of 400 degrees. At °C, the impurities in the sample are removed by calcination, after which we obtain pure zinc oxide nanoparticles (Nayak et al.,2020).

### **2.4-The mechanism of action of nanoparticles to inhibit toxins or toxin-producing fungi**

The most important thing that distinguishes nanoparticles is their extremely small size, as they work to penetrate cell membranes, which leads to their damage, that is, they deliberately form holes on the surface of the cells, thus accumulating them and leading to cell death due to the disturbances occurring in the fungal hyphae as a result of the effect of nanoparticles on the sugar and protein content in the fungal hyphae (Jogee.,2020; Prabhu & Poulouse.,2012; Ouda.,2014).

### **2.5-Isolation and identification of mycotoxin-producing fungi**

A portion of the samples is taken and placed on PDA medium containing 0.05 mg of Tetracycline and incubated at a temperature of 25 degrees Celsius for a period ranging from one to five days, after which we observe the growth of different types of fungi that are diagnosed according to their morphological appearance in terms of the shape of the colonies, conidia, and pods according to ( Atef et al.,2019; Pitt & Hocking.,2009).

### **2.6- Using zinc oxide nanoparticles at different concentrations on some types of toxin-producing fungi.**

Several concentrations (550 µg/mL. 600 µg/mL. 650 µg/mL. 700 µg/mL. 750 µg/mL. 800 µg/mL) of previously prepared zinc oxide nanoparticles were used, with distilled water added to them to make a diluent. After that, three replicates were made for each concentration, and 1 ml of each concentration was taken and added before pouring the PDA medium into the pouring dishes. After pouring the medium and solidifying, the plates were inoculated with the isolated fungi, and Tetracycline antibiotic was used on some of the plates as a positive control. Then the plates were left

untreated and containing the fungal samples as a negative control, The incubation process was carried out at a temperature of 25 degrees Celsius for a period of (1-5) days.

## Results and Discussion

### 3.1-Isolation and diagnosis of some types of fungi that produce mycotoxins

The table below shows the number of samples taken for the purpose of the study that were grown on PDA medium containing 0.05 mg tetracycline/ml for a week and at an appropriate temperature. 15 samples were taken from corn fruits, so the number of infected samples was only two, i.e. 13.3 of the total number. As for peanuts, approximately 50 samples were taken, with the number of samples being 23 infected, or 46%. As for vegetable samples, only 7 samples were taken from onions, so the number of samples infected with the fungus was 3, meaning 42%. As for tomatoes, the number of samples taken was 20, while 15 infected samples were taken, i.e. 75%. As for sweet potatoes, it was The number of samples taken was 13, while the infected samples were 7, meaning a percentage of 53.8%. As for grapes, the number of samples taken was 23, while the infected samples were 19, meaning a percentage of 82.6%. As for watermelon, the number of samples taken was two, while the samples were infected with both samples, meaning a percentage of 100%. As for juices, they were a number The samples taken were only three samples, while the infected samples were one sample, meaning a percentage of 33.3%. As for milk, the number of samples taken was 7 samples, while the infected samples were one sample, meaning a percentage of 14.2%. Finally, bread, where the number of samples taken was 9 samples, while the number of infected samples was 4 samples, meaning a percentage of 44.4%. It was isolated and diagnosed based on some morphological characteristics.

**Table (1) shows Number of isolated samples**

No	Sample	Number	Infected sample	Intact sample	Infection rate
1	Corn	15	2	13	% 13.3
2	Peanut	50	23	27	% 46
3	Onion	7	3	4	% 42
4	Tomato	20	15	5	% 75
5	Sweet potato	13	7	6	% 53.8
6	grapes	23	19	4	% 82.6
7	Watermelon	2	2	0	% 100
8	Fruit juices	3	1	2	% 33.3
9	milk	7	1	6	% 14.2
10	Bread	9	4	5	% 44.4

The table below shows the type of fungi obtained from the different samples, Growth of the *Aspergillus* sp fungus appeared in samples taken from corn, peanuts, onions, juices, grapes, and milk, This type of fungus is capable of producing aflatoxin, and this is consistent with (El-Hamaky et al., 2023). While *Fusarium* sp fungi were obtained from tomato, sweet potato and watermelon samples. *Fusarium* sp. was found in tomatoes, sweet potatoes, and watermelon, *Penicillium* sp was found in Bread, as the most important characteristic of this type of fungus is that it produces several toxins such as Aflatoxin, Ochratoxins, Patulin, Oxalic acid, Malformin, Fumonisin, Zearalenone, as well as Penicillin.

### 3.2-Types of fungi and the toxins they produce.

**Table (2) shows some types of fungi and the toxins they produce**

No	Sample	Mycotoxins secreting fungi	Mycotoxins
1	Corn	A. flavus	Aflatoxin
2	Peanut	Aspergillus sp	Aflatoxin
3	Onion	Aspergillus niger	Ochratoxins
4	Fruit juices	Aspergillus sp, Penicillium sp	Ochratoxins
5	Grapes	A.niger	Fumonisin
6	Milk	Aspergillus niger	Aflatoxin
7	Tomato	F. oxysporum	Zearalenone
8	Sweet potato	F. oxysporum	Zearalenone
9	Watermelon	Fusarium sp	Zearalenone
10	Bread	Penicillium sp	Penicillin

### 3.3- Using zinc oxide nanoparticles and testing their effect on some types of toxin-producing fungi.

We are working on cultivating the fungi that were isolated from some samples and obtained in their pure form and then treating them with previously prepared nanoparticles and observing the effect of these particles on the growth of the fungus at different concentrations.

**Table (3) Shows zinc oxide nanoparticles and their effect on some harmful fungi**

	A. flavus	A. niger	F. oxysporum	Penicillium sp
<b>Con.550 µg/mL</b>	3.6	3.3	3.8	4.1
<b>Con.600 µg/mL</b>	3	2.9	2.8	3.6
<b>Con.650 µg/mL</b>	2.5	2.5	2	3
<b>Con.700 µg/mL</b>	1.8	1	0.9	2.1
<b>Con.750 µg/mL</b>	0.5	0.2	0	0.8
<b>Con.800 µg/mL</b>	0	0	0	0
<b>tetracycline</b>	1.9	1	0.5	0.7

The effect of zinc oxide nanoparticles on a few isolated toxin-producing fungi, such as A. flavus, is demonstrated by the study results in Table 3. The average diameter of the radial growth inhibition zone reached 3.6 when it was applied at a dose of 550 µg/ml. This prevented the development of mycotoxins, but to a small extent compared to the other concentrations. The average diameter of the radial growth inhibition zone reached three when used at a concentration of 600 µg/ml. The average diameter of the radial growth inhibition zone was 2.5 when applied at a concentration of 650 µg/ml. The average diameter of the radial growth inhibition zone was 1.8 at a concentration of 700 µg/ml, and 0.5 at a concentration of 750 µg/ml. The plates treated with the tetracycline antibiotic as a positive control had an average diameter of 1.9, while the zone of radial growth inhibition at a dose of 800 µg/ml had an average diameter of 0. Compared with the other concentrations and the tetracycline antibiotic, the average diameter of the radial growth inhibition zone for the concentration was 800. It is the best and largest in terms of inhibition.

When these concentrations were applied to the fungus A. niger, it became clear from the results of the study that the average diameter of the inhibition zones for radial growth reached (3.3,



2.9, 2.5, 1, 0.2, 0), respectively, when applied at the following doses (550, 600, 650, 700, 750, 800) micrograms/ml, respectively, while when adding the tetracycline antibiotic to the media, the rate of inhibition of the fungus was 1. We note that at the concentration of 800, it was better and greater in terms of inhibition.

As for the *F. oxysporum* fungus, applying these concentrations we notice different results, as the zinc nanoparticles had a clear inhibitory effect on the *F. oxysporum* fungus. It was clear from the results of the study that the average diameter of the inhibition zones for radial growth reached (3.8, 2.8, 2, 0.9, 0.9, 0), respectively, when applied to the following doses (550, 600, 650, 700, 750) micrograms/ml on respectively, while when the antibiotic tetracycline was added to the medium, the fungal inhibition rate was 0.5 as a positive control. We note that at a concentration of 750, it was better and greater in terms of inhibition.

Finally, we apply and add these concentrations to the dishes that will be inoculated with the *Penicillium* sp fungus and monitor the inhibitory rates of the nanoparticles. The results of the study showed that the average diameter of the inhibition zones for radial growth reached (4.1, 3.6, 3, 2.1, 0.8, 0), respectively, when applied to the following doses (550, 600, 650, 700, 750, 800) micrograms/ml. Respectively, when adding the antibiotic tetracycline to the medium, the fungal inhibition rate was 0.7 as a positive control factor. We note that at a concentration of 800, it was better and greater in terms of inhibition and thus preventing the secretion of mycotoxins.

These results were compared with the results of dishes that were not treated with any concentration of antibody as a negative control agent, where the average diameter of the radial growth inhibition zone was 7.5.

## Conclusion

This study, as well as previous studies, propose the use of zinc oxide nanoparticles, which have antibacterial activity on several types of toxin-producing fungi, and which are considered a promising alternative to many antibiotics that have been proven to be resistant to organisms. Nanotechnology is also considered one of the most effective technologies in Combating bacteria and fungi. These are considered promising ways to prevent the growth of fungi and eliminate their harmful and dangerous effects.

## References

- [1] Aiad, I., El-Sukkary, M. M., Soliman, E. A., El-Awady, M. Y., and Shaban, S. M., "In Situ and Green Synthesis of Silver Nanoparticles and Their Biological Activity," *Journal of Industrial and Engineering Chemistry*, vol. 20, no. 5, pp. 3430-3439, 2014.
- [2] Ashour, E. A., El-Hack, M. E. A., Shafi, M. E., Alghamdi, W. Y., Taha, A. E., Swelum, A. A., and El-Saadony, M. T., "Impacts of Green Coffee Powder Supplementation on Growth Performance, Carcass Characteristics, Blood Indices, Meat Quality and Gut Microbial Load in Broilers," *Agriculture*, vol. 10, no. 10, p. 457, 2020.
- [3] Aslam, N., Rodrigues, I., McGill, D. M., Warriach, H. M., Cowling, A., Haque, A., and Wynn, P. C., "Transfer of Aflatoxins from Naturally Contaminated Feed to Milk of Nili-Ravi Buffaloes Fed a Mycotoxin Binder," *Animal Production Science*, vol. 56, no. 10, pp. 1637-1642, 2015.
- [4] Atef, A. H., Noha, H. O., and Manal, M., "Detection of Mycotoxigenic *Fusarium* Species in Poultry Rations and Their Growth Control by Zinc Nanoparticles," *Journal of Environmental Science and Technology*, vol. 15, no. 4, pp. 327-338, 2021.
- [5] Cheat, S., Oswald, I. P., and Kolf-Clauw, M., "Mycotoxin Outbreak in Animal Feed," in

- Foodborne Diseases Case Studies of Outbreaks in the Agri-Food Industries*, pp. 411, 2016.
- [6] Cunha, S. C., Sa, S. V., and Fernandes, J. O., "Multiple Mycotoxin Analysis in Nut Products: Occurrence and Risk Characterization," *Food and Chemical Toxicology*, vol. 114, pp. 260-269, 2018.
- [7] Dellaflora, L., Dall'Asta, C., and Galaverna, G., "Toxicodynamics of Mycotoxins in the Framework of Food Risk Assessment—An In Silico Perspective," *Toxins*, vol. 10, no. 2, p. 52, 2018.
- [8] El-Hamaky, A. M., Hassan, A. A., El Yazeed, H. A., and Refai, M. K., "Prevalence and Detection of Toxigenic *A. flavus*, *A. niger*, and *A. ochraceus* by Traditional and Molecular Biology Methods in Feeds," *International Journal of Current Research*, vol. 8, no. 1, pp. 25621-25633, 2016.
- [9] El-Hamaky, A. M., Hassan, A. A., Wahba, A. K., and El-Mosalamy, M. M., "Influence of Copper and Zinc Nanoparticles on Genotyping Characterizations of Multi-Drug Resistance Genes for Some Calf Pathogens," *International Journal of Veterinary Science and Research*, vol. 9, no. 2, pp. 90-99, 2023.
- [10] Elkenany, R., and Awad, A., "Types of Mycotoxins and Different Approaches Used for Their Detection in Foodstuffs," *Mansoura Veterinary Medical Journal*, vol. 22, no. 1, pp. 25-32, 2021.
- [11] El-Sayed, R. A., Jebur, A. B., Kang, W., and El-Demerdash, F. M., "An Overview on the Major Mycotoxins in Food Products: Characteristics, Toxicity, and Analysis," *Journal of Future Foods*, vol. 2, no. 2, pp. 91-102, 2022.
- [12] El-Tarabily, K. A., El-Saadony, M. T., Alagawany, M., Arif, M., Batiha, G. E., Khafaga, A. F., and Abd El-Hack, M. E., "Using Essential Oils to Overcome Bacterial Biofilm Formation and Their Antimicrobial Resistance," *Saudi Journal of Biological Sciences*, vol. 28, no. 9, pp. 5145-5156, 2021.
- [13] Freire, L., and Sant'Ana, A. S., "Modified Mycotoxins: An Updated Review on Their Formation, Detection, Occurrence, and Toxic Effects," *Food and Chemical Toxicology*, vol. 111, pp. 189-205, 2018.
- [14] Gacem, M. A., Gacem, H., Telli, A., and Khelil, A. O. E. H., "Mycotoxins: Decontamination and Nanocontrol Methods," in *Nanomycotoxicology*, Academic Press, pp. 189-216, 2020.
- [15] Geethalakshmi, R., and Sarada, D. V. L., "Characterization and Antimicrobial Activity of Gold and Silver Nanoparticles Synthesized Using Saponin Isolated from *Trianthema Decandra* L.," *Industrial Crops and Products*, vol. 51, pp. 107-115, 2013.
- [16] Horky, P., Skalickova, S., Baholet, D., and Skladanka, J., "Nanoparticles as a Solution for Eliminating the Risk of Mycotoxins," *Nanomaterials*, vol. 8, no. 9, p. 727, 2018.
- [17] Jayachandran, A., Aswathy, T. R., and Nair, A. S., "Green Synthesis and Characterization of Zinc Oxide Nanoparticles Using *Cayratia Pedata* Leaf Extract," *Biochemistry and Biophysics Reports*, vol. 26, p. 100995, 2021.
- [18] Jiang, K., Huang, Q., Fan, K., Wu, L., Nie, D., Guo, W., and Han, Z., "Reduced Graphene Oxide and Gold Nanoparticle Composite-Based Solid-Phase Extraction Coupled with Ultra-High-Performance Liquid Chromatography-Tandem Mass Spectrometry for the Determination of 9 Mycotoxins in Milk," *Food Chemistry*, vol. 264, pp. 218-225, 2018.
- [19] Jogee, P., and Rai, M., "Application of Nanoparticles in Inhibition of Mycotoxin-Producing Fungi," in *Nanomycotoxicology*, Academic Press, pp. 239-250, 2020.

- [20] Kołodziejczak-Radzimska, A., and Jesionowski, T., "Zinc Oxide—from Synthesis to Application: A Review," *Materials*, vol. 7, no. 4, pp. 2833-2881, 2014.
- [21] Krstanovic, V., Šarkanj, B., Velic, N., Mastanjevic, K., Šantek, B., and Mastanjevic, K., "Mycotoxins in Malting and Brewing By-Products Used for Animal Feed," in *European Biotechnology Congress 2017*, pp. S68-S69, 2017.
- [22] Moodley, J. S., Krishna, S. B. N., Pillay, K., Serphen, F., and Govender, P., "Green Synthesis of Silver Nanoparticles from Moringa Oleifera Leaf Extracts and Its Antimicrobial Potential," *Advances in Natural Sciences: Nanoscience and Nanotechnology*, vol. 9, no. 1, p. 015011, 2018.
- [23] Mukunthan, K. S., and Balaji, S., "Cashew Apple Juice (Anacardium Occidentale L.) Speeds Up the Synthesis of Silver Nanoparticles," *International Journal of Green Nanotechnology*, vol. 4, no. 2, pp. 71-79, 2012.
- [24] Nayak, R., Ali, F. A., Mishra, D. K., Ray, D., Aswal, V. K., Sahoo, S. K., and Nanda, B., "Fabrication of CuO Nanoparticle: An Efficient Catalyst Utilized for Sensing and Degradation of Phenol," *Journal of Materials Research and Technology*, vol. 9, no. 5, pp. 11045-11059, 2020.
- [25] Ouda, S. M., "Antifungal Activity of Silver and Copper Nanoparticles on Two Plant Pathogens, Alternaria Alternata and Botrytis Cinerea," *Research Journal of Microbiology*, vol. 9, no. 1, pp. 34-40, 2014.
- [26] Pelaz, B., Jaber, S., De Aberasturi, D. J., Wulf, V., Aida, T., de la Fuente, J. M., and Parak, W. J., "The State of Nanop